

Preparation of Ornamental Fish for Shipping¹

Tina C. Crosby, Jeffrey E. Hill, Carlos V. Martinez, Craig A. Watson, Deborah B. Pouder, and Roy P.E. Yanong²

Introduction

Handling of ornamental fish from the time they are harvested through shipment affects their overall quality. Quality of marketable fish is directly related to their health, which is affected by the amount of stress and trauma they experience during harvest, holding, and transport (see IFAS Fact Sheets FA-117 *Harvesting Ornamental Fish From Ponds*, FA-118 *Grading Ornamental Fish*, FA-119 *On-Farm Transport of Ornamental Fish*, and FA-3 *Fish Handling and Transport*). Stress weakens the immune system of the fish, resulting in increased susceptibility to disease (see IFAS Circular 919 *Stress – Its Role in Fish Disease*). There is no substitute for careful handling and good water quality when trying to optimize the health of the fish. As with the other steps in the harvesting, grading, and transport of fish, proper care and planning when preparing ornamental fish for shipping leads to a healthier and more marketable product.

Holding Fish Prior to Shipping

In a typical Florida production setting, harvested ornamental fish are brought in from ponds and placed into a holding facility, where they are often held for several days prior to shipping (see IFAS Fact Sheet FA-117 *Harvesting Ornamental Fish from Ponds*). During this time, the fish are allowed to clear out their digestive tracts, which reduces the amount of waste released into transport bags and helps maintain acceptable water quality during shipping.

Fish should be examined for appearance (e.g., ragged fins, ulcers, fungal patches) and behavior upon arrival at the holding facility (see IFAS Fact Sheet FA-118 *Grading Ornamental Fish*). If possible, fish should also be checked for microscopic parasites by analyzing biopsied tissues from gill, skin, and fins especially if there are external signs of disease such as white spots, excess mucus, visible scale loss, or abnormal behavior. Consult a fish health specialist for assistance.

1. This document is FA-120, one of a series of the Fisheries and Aquatic Sciences Department, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida. Original publication date July 2005. Revised December 2007. Reviewed January 2011. Visit the EDIS website at <http://edis.ifas.ufl.edu>.

2. Tina C. Crosby, Graduate Assistant; Jeffrey E. Hill, Assistant Professor; Carlos V. Martinez, Assistant In Extension; Craig A. Watson, Research Coordinator, Deborah B. Pouder, Coordinator of Research Programs and Services, and Roy P.E. Yanong, Associate Professor; Tropical Aquaculture Laboratory, 1408 24th St. SE, Ruskin, FL 33570, Department of Fisheries and Aquatic Sciences, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida, Gainesville, FL 32611

The use of trade names in this publication is solely for the purpose of providing specific information. UF/IFAS does not guarantee or warranty the products named, and references to them in this publication does not signify our approval to the exclusion of other products of suitable composition. All chemicals should be used in accordance with directions on the manufacturer's label.

The Institute of Food and Agricultural Sciences (IFAS) is an Equal Opportunity Institution authorized to provide research, educational information and other services only to individuals and institutions that function with non-discrimination with respect to race, creed, color, religion, age, disability, sex, sexual orientation, marital status, national origin, political opinions or affiliations. U.S. Department of Agriculture, Cooperative Extension Service, University of Florida, IFAS, Florida A. & M. University Cooperative Extension Program, and Boards of County Commissioners Cooperating. Millie Ferrer-Chancy, Interim Dean

Prophylactic Treatments

There are very few chemicals that are approved by the Food and Drug Administration (FDA) for use in aquaculture. In fact, there are few to no chemicals officially approved by the FDA for use in ornamental fish. In order to assist the ornamental fish industry and allow access to certain unapproved products until this situation is reversed, the FDA has chosen to exercise regulatory discretion. If there is a significant threat to human health or the environment, the FDA may prosecute for illegal use. Oxytetracycline is approved for use in catfish to control bacterial hemorrhagic septicemia and pseudomonas disease. Oxytetracycline is approved for use in a few other species. See the FDA-CVM Aquaculture website for additional information.

FDA-approved formalin products that control external protozoa (e.g., *Chilodonella*, *Ichthyobodo*, *Epistylis*, *Trichodina*, *Ichthyophthirius*, monogenetic trematodes) of finfish and/or fungi (i.e., family Saprolegniaceae) on finfish eggs are available from only three companies: Natchez Animal Supply Company (Formalin-F®), Argent Laboratories (Paracide-F®), and Western Chemical (Parasite-S®).

Salt (NaCl) is considered unapproved but is of low regulatory priority. Low regulatory priority is not considered an approval or affirmation of safety or effectiveness, but if used as a 0.5%-1.0% solution as an osmoregulatory aid or a 3% solution for 10-30 minutes as a parasiticide, the FDA is unlikely to object to its use.

Potassium permanganate is currently unapproved, but is in a special category—regulatory action is deferred pending further study. Acriflavine neutral, methylene blue, and nitrofurazone are not approved and are of high enforcement priority if used illegally in foodfish and use is subject to regulatory action on a case-by-case basis. Malachite green used in Quick Cure® is also a high enforcement priority chemical if used illegally in foodfish.

The use of other approved animal and human drugs is allowed under supervision of a veterinarian providing that all requirements of the drugs'

regulation are met and the drug is not administered through feed. However, nontherapeutic use of a drug is not permitted. Additional information regarding FDA drug approval can be found at the FDA-CVM Aquaculture website and in IFAS Circular 84 *Use of Antibiotics in Ornamental Fish Aquaculture*.

If fish are generally healthy with no history of problems, use of a prophylactic treatment (Figure 1) may be an unnecessary cost. Furthermore, unnecessary prophylactic chemical treatments can do more harm than good. Excessive or incorrect use of a prophylactic treatment can increase resistance to the chemical and decrease its effectiveness. However, in some instances, prior to shipping, prophylactic treatment with a chemical bath or dip may help reduce external parasites and bacterial loads. The use of a prophylactic or preventative treatment may decrease the outbreaks of disease while fish are recovering from harvesting and handling and increase their survival during shipment.

Some chemicals (e.g., potassium permanganate, formalin) may damage fish gills or skin and should be used carefully and only when absolutely necessary. Chemicals may also adversely affect water quality. For example, formalin decreases dissolved oxygen concentrations in water. Ideally, the producer should be familiar with common health problems, actively work to identify and treat them as they become apparent, and seek assistance from a fish health professional when needed.



Figure 1. Treatment Chemicals Credits: Tina Crosby 2004

Experiments evaluating eight common chemicals used for prophylaxis on fish prior to shipment were conducted at the University of Florida Tropical Aquaculture Laboratory. The chemicals tested were oxytetracycline, formalin, salt (3ppt sodium chloride, NaCl), potassium permanganate (KMnO₄),

acriflavine neutral, methylene blue, nitrofurazone, and Quick Cure® (a mixture of 99.2% formalin and 0.75% malachite green mix) (Table 1). Fish were trapped in a pond, graded, and placed into holding tanks with flow-through aerated well water. Each chemical was applied according to the industry standard to separate tanks as 11.5-hour baths, except for potassium permanganate, which was administered as a 4-hour bath.

Cure®, or formalin significantly improved the overall appearance and behavior of fish over time compared to no treatment. In contrast, fish treated with acriflavine neutral were inferior to fish not treated with any chemical in both appearance and behavior. Methylene blue and salt did not have any effect on the appearance or behavior of the fish when used as a prophylactic bath.

Table 1. Prophylactic treatments tested

Treatment	Usage	Dosage	Time in Bath	Overall Appearance	Overall Effectiveness	FDA-Approved for use in aquaculture
Salt	Aid Osmoregulation	3 ppt	11.5 hours	None	None	No
Acriflavine neutral	Antimicrobial - (Anti-microscopic organisms)	7 mg/L	11.5 hours	Bad	Bad	No
Formalin (37%)	Antiparasitic - (ecto)	1 ml/10 gal	11.5 hours	Good	Good	Yes
Methylene Blue	Antiparasitic - (ecto) Antibacterial	2 mg/L	11.5 hours	None	None	No
Nitrofurazone	Antibacterial	10 mg/L	11.5 hours	Good	Good	No
Potassium Permanganate	Antiparasitic - (ecto) Antimicrobial	2 mg/L	4 hours	Better	Better	No
Quick Cure (99.2% formalin + 0.75% malachite green mix)	Antiparasitic - (ecto); esp. <i>Ichthyophthirius multifiliis</i> ("Ich") Antifungal	0.5 ml/10 gal	11.5 hours	Better	Better	No
Oxytetracycline	Antibacterial	1 g/10 gal	11.5 hours	Better	Better	Yes

Following treatment, fish were observed for one-week. Fish were evaluated for loss of scales, torn fins, coloration, deformities, pop eye, parasites, and hemorrhaging. Fish behavior was also evaluated for abnormal signs including spinning, flashing, clamping, hanging, piping, gasping, as well as activity level. Appearance and behavior for each tank of fish were rated on a scale of 1-5, with 1 being overall unfit for sale (problems with >50% of fish) to 5 being superior quality. The use of tetracycline, potassium permanganate, nitrofurazone, Quick

Oxytetracycline is an antibiotic, used to treat bacterial infections. The positive results seen with the use of oxytetracycline in our experiments may have been due to the presence of susceptible bacteria. Oxytetracycline may not be as effective under other conditions. However, in general, a single dose treatment of oxytetracycline is not recommended. Additionally, overuse or misuse of oxytetracycline or any other antibiotic, may lead to bacterial resistance making the drug ineffective. Before using any antibiotic, confirmation of a bacterial infection should

be established and proper regimes should be followed (see IFAS Circular 84 *Use of Antibiotics in Ornamental Fish Aquaculture*). Note that oxytetracycline binds to calcium ions in the water, resulting in the need for a higher dosage of oxytetracycline in hard water [Total Hardness (TH) > 120 ppm]. Also keep in mind the legal issues for use of any chemical.

Shipping to a Buyer

Once the fish have had time (1-3 days) to evacuate the wastes in their digestive tracts and have been properly evaluated and treated for any parasites or disease, they are ready to be shipped to a buyer. Fish should be placed into plastic bags containing oxygenated water and put into a polystyrene shipping box. The polystyrene boxes are placed into a cardboard box for shipping. It is important to use shipping water that has water quality appropriate for the fish being shipped (e.g., pH, temperature, TAN, NO₂, DO) (see IFAS Fact Sheet FA-119 *On Farm Transport of Ornamental Fish*). Water quality in the bag will become degraded due to accumulation of carbon dioxide, ammonia, and other waste products during shipment.

Temperature

Temperature is an important consideration when transporting fish. Different species of fish thrive at different temperatures: tropical fish do better at temperatures 26°C - 30°C (> 78.8°F - 86°F), warm water fish thrive at temperatures 21°C - 28°C (69.8°F - 82.4°F), and cool water fish do best at temperatures 7 - 15°C (44.6°F - 59°F) (Noga, 2000, Avault 1996, Wedemeyer 1996). During the summer and winter months, or when transport times are long, ice gel packs or heat packs can be placed into shipping boxes to help maintain an appropriate water temperature. The ideal temperature for the shipping water is dependent on the species of fish being transported.

Shipping Additives

Chemical additives are often used in transport water to sedate the fish or reduce pathogen (any agent that causes disease) loads in the water (Figure 2).

Experiments at the University of Florida Tropical Aquaculture Laboratory were run to test various shipping chemical additives for their effects on appearance and behavior of fish post shipping. The chemicals tested were: tricaine methanesulfonate (MS-222), salt (3ppt NaCl), methylene blue, acriflavine neutral, quinaldine, eugenol, an experimental commercial product in the form of a feed, and a water additive.

The commercial product contained arachidonic acid (ArA), a fungus extract from *Mortierella sp.*, claimed by the manufacturer to be used to regulate immune systems; and an ethanol extract from the genus *Opuntia*, a cactus species, claimed by the manufacturer to be used to stimulate the production of heat shock proteins (HSPs) which work to counteract stress responses.

Tricaine methanesulfonate is approved for use in finfish as a sedative and anesthetic. Quinaldine and eugenol are not approved for use in aquaculture. A description of approval for the other chemicals tested in this experiment were described in the "Prophylactic Treatments" section of this publication.



Figure 2. Shipping additives [tricaine methanesulfonate, sodium thiosulfate, salt (NaCl)] and supplies Credits: Tina Crosby 2004

Velvet swordtail fish were trapped from a pond and placed into holding tanks with flow-through aerated well water for four days. The fish receiving the commercial test feed were fed for three days prior to shipping. The day of transportation, fish were transported with one of the additives added to the shipping water (1.5L) (Table 2). The fish were transported by van for two hours in 1/4 square-bottom shipping bags.

Fish appearance and behavior were evaluated after harvest, immediately prior to transport, and one week after transport. Immediately following transport, fish treated with methylene blue, salt, MS-222, quinaldine, eugenol, the experimental water additive, and control fish all had better appearance than those treated with acriflavine neutral. Fish, treated with methylene blue or salt, had the best appearance.

After one week of holding, fish from all treatment groups appeared the same. Fish treated with methylene blue or salt had a better appearance than control fish post transport. Fish, treated with salt or MS-222, had better behavior following the one-week post transportation holding period. Acriflavine neutral had a detrimental effect upon the appearance and behavior of fish.

Another set of experiments (T.M. Corona, J.E. Hill, C.A. Watson, and R.P.E. Yanong, 2006; manuscript in prep) investigated the efficacy of acriflavine (8 mg/L), methylene blue (2 mg/L), nitrofurazone (4 mg/L), and two commercial products (Blue Holdex® (125 mg/L), Jungle Laboratories; and Furazone Green (26mg/L), First Choice™) as transport additives for control of *Aeromonas hydrophila* and other culturable aquatic bacteria in shipping bags of ornamental tropical fish. The dosage rates tested were industry standards.

Experiment 1 consisted of simulated shipping field trials using swordtails *Xiphophorus hellerii*. One of the five shipping compounds was added to test effects on bacterial loads as a whole during transport compared to untreated controls. Experiment 2 was run to determine the effects of these shipping chemicals against one particular clinical strain of *Aeromonas hydrophila*. *Aeromonas hydrophila* was added to sterilized well water in shipping bags containing no fish. Next, one of the five shipping additives was added to the shipping bags to examine its effect on that species and strain of bacteria. In both trials, the water containing the bacteria was sampled every eight hours to determine how effective each of the shipping additives was in killing aquatic bacteria over time.

Data on the mean number of bacteria obtained at time 0, 8, 16, and 24hr were assessed for each of the five separate shipping additives as well as the control.

In experiment 1, overall bacterial populations increased over the 24 hours of the experiment in each of the water samples obtained at time 8, 16, 24hr. There was less of an increase at 8 hours for acriflavine, but by 24 hours, there was no significant difference between any of the treatments and the control. This suggests that none of the treatments had greater efficacy over the others by the end of the 24 hours for total culturable bacteria populations. Results from experiment 2, testing only one clinical strain of the bacteria *Aeromonas hydrophila* showed there was no obvious pattern of increase in bacteria populations over the 24 hours during the experiment. At different time points, some treatments had increases in bacterial numbers, while others had decreases. However, overall, acriflavine did depress *Aeromonas* numbers at times 8, 16, and 24hr significantly, relative to control and all other treatments.

While acriflavine kept *Aeromonas* bacteria numbers lower than other treatments in the experiment with no fish and only bacteria, in experiment 2, with fish in transport, acriflavine did not appear to provide any long-term bactericidal properties. Because of these results evaluating the appearance and behavior of fish transported in acriflavine and the lack of long-term bactericidal effects, acriflavine is not recommended for use.

Shipping Bags and Boxes

Square-bottom bags are ideal for shipping fish because they sit upright in shipping boxes and do not have “folds” where fish can concentrate and become trapped (Figure 3 and Figure 4). Another benefit of the square-bottom bag is that it allows for a larger surface area between the oxygen and the water, maximizing oxygen diffusion into the water reducing localized zones of oxygen depletion. The size of the bag that is used depends on the number, size, and species of the fish being shipped. Table 3 lists dimensions of bags commonly used to ship

Table 2. Shipping additives tested

Treatment	Usage	Dosage	Overall Appearance	Overall Behavior	FDA-Approved for use in Aquaculture
Salt	Aid osmoregulation	3 ppt	Good	Good	No
Acriflavine neutral	Antimicrobial	7 mg/L	Bad	Bad	No
Methylene Blue	Antimicrobial	2 mg/L	Good	Good	No
Eugenol	Sedative	5 mg/L	No Effect	No Effect	No
MS-222	Sedative	20 mg/L	No Effect	Good	Yes
Quinaldine	Sedative	5 mg/L	No Effect	No Effect	No
Experimental Feed Additive	Lower Stress Response	3 days fed before shipping	No Effect	No Effect	No
Experimental Water Additive	Lower Stress Response	0.5 ml/L	No Effect	No Effect	No

ornamental fish for prolonged periods of time (up to 36 hours). Table 4 lists common species, sizes, and numbers of fish shipped from Florida in square-bottom bags with the standard 2 gallons of high-quality transport water (table is located at the end of this publication). In 1/4 and 1/2 square-bottom shipping bags, the oxygen gas volume to water volume ratio is generally 60% oxygen to 40% water.

Pillow bags do not have square bottoms, so they lie on their sides. They are less expansive than square-bottom bags and come in a variety of sizes.

If a shipping additive is to be used, it is placed into the bag water prior to the addition of fish. Once the fish are placed into the appropriate bag, the ambient air is removed (squeezed out) and replaced with pure oxygen from an oxygen bottle. The bag is then sealed with a rubber band or other sealing device. During transport, oxygen levels decrease and carbon dioxide levels increase as fish respire. The addition of pure oxygen to the shipping bag allows the producer to ship a greater number of fish in each bag for a longer period of time. If the sealed bag contains only ambient air, the fish quickly consume

all of the available oxygen resulting in high mortalities (see IFAS Fact Sheet FA-119 *On Farm Transport of Ornamental Fish*).

There are a variety of shipping box sizes available such as the standard "double" box that holds two full bags of fish. The size of box is determined by the number of fish being shipped. Once prepared, bags destined for local transport are put into the Florida standard ornamental "single" polystyrene boxes (17-inch x 17-inch) with lids. If fish are going further than a local buyer, the polystyrene boxes are sealed with packing tape and placed into tight-fitting cardboard outer boxes (17 1/4-inch x 17 1/4-inch) that provide added protection during shipment. Boxes are usually pre-labeled with a "live fish" stamp and are suitable for shipping via airplane or truck (Figure 5).

Permits and Health Certificates

Permit requirements and health certificates vary by state and region. Check with your local state

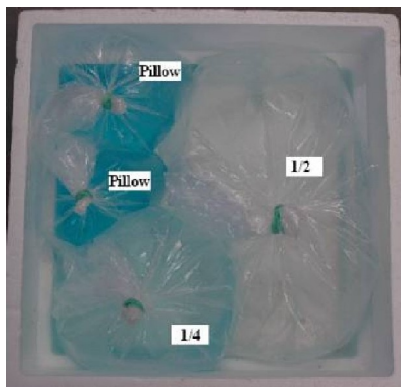


Figure 3. Pillow and square-bottom shipping bags in box
Credits: Tina Crosby 2004



Figure 4. Full square-bottom shipping bag Credits: Tina Crosby 2004

Table 3. Dimensions for bags commonly used to ship ornamental fish from Florida

Bag Size	Length x Width x Height
Full Square-Bottom Bags	15.5 inch x 14.5 inch x 22.5 inch
1/2 Square-Bottom Bags	16 inch x 8 inch x 22 inch
1/4 Square-Bottom Bags	7.5 inch x 6.5 inch x 20.5 inch
Pillow Bags	Various (10 inch x 20 inch pictured in Figure 3)



Figure 5. Moving fish with a fork-lift at the airport between planes Credits: Jon Kao 2004

agency and/or the importing country's regulations to determine requirements for shipping ornamental fish domestically and internationally.

Summary

The quality and overall health of ornamental fish are affected by stressors that occur during harvest, holding, and transport to the buyer. Experienced ornamental fish farmers minimize the effects of these stressors. Overall fish health is improved by screening fish showing signs of disease. If harvested fish show signs of external parasites, the prophylactic use of potassium permanganate, Quick Cure®, or formalin may provide protection from mortality. The use of salt (3ppt) or methylene blue in shipping water may be beneficial to the overall behavior and appearance of fish post-transportation. Acriflavine neutral is not recommended for use because it has a negative effect on the appearance and behavior of fish. Optimal dissolved oxygen and temperature levels should be maintained during shipping. Prior to shipment, determine the best packaging methods (e.g., bag size, amount of water per bag, amount of fish per bag, addition of ice or heat pack). These factors, along with good water quality, help facilitate the production and sale of high quality marketable fish.

Recommended Reading

UF/IFAS Circular 84 *Use of Antibiotics in Ornamental Fish Aquaculture*.
<http://edis.ifas.ufl.edu/FA084>

UF/IFAS Circular 120 *Fish Health Management Considerations in Recirculating Aquaculture Systems - Part 1: Introduction and General Principles*.
<http://edis.ifas.ufl.edu/FA099>

UF/IFAS Circular 121 *Fish Health Management Considerations in Recirculating Aquaculture Systems - Part 2: Pathogens*. <http://edis.ifas.ufl.edu/FA100>

UF/IFAS Circular 122 *Fish Health Management Considerations in Recirculating Aquaculture systems - Part 3: General Recommendations and Problem-Solving Approaches*.
<http://edis.ifas.ufl.edu/FA101>

UF/IFAS Circular 716 *Introduction to Freshwater Fish Parasites*.
<http://edis.ifas.ufl.edu/FA041>

UF/IFAS Circular 919 *Stress - Its Role in Fish Disease*. <http://edis.ifas.ufl.edu/FA005>

UF/IFAS Fact Sheet FA-3 *Fish Handling and Transport*. <http://edis.ifas.ufl.edu/FA019>

UF/IFAS Fact Sheet FA-7 *Fish Fingerlings: Purchasing, Transporting, and Stocking*.
<http://edis.ifas.ufl.edu/FA013>

UF/IFAS Fact Sheet FA-12 *Principles of Water Recirculation and Filtration in Aquaculture*.
<http://edis.ifas.ufl.edu/FA050>

UF/IFAS Fact Sheet FA-13 *Use of Copper in Freshwater Aquaculture and Farm Ponds*.
<http://edis.ifas.ufl.edu/FA008>

UF/IFAS Fact Sheet FA-16 *Ammonia*.
<http://edis.ifas.ufl.edu/FA031>

UF/IFAS Fact Sheet FA-27 *Dissolved Oxygen for Fish Production*.
<http://edis.ifas.ufl.edu/FA002>

UF/IFAS Fact Sheet FA-29 *Introduction to Viral Diseases of Fish*. <http://edis.ifas.ufl.edu/FA034>

UF/IFAS Fact Sheet FA-37 *Use of Potassium Permanganate to Control External Infections of Ornamental Fish*. <http://edis.ifas.ufl.edu/FA027>

UF/IFAS Fact Sheet FA-55 *Submission of Fish for Diagnostic Evaluation*.
<http://edis.ifas.ufl.edu/FA055>

UF/IFAS Fact Sheet FA-117 *Harvesting Ornamental Fish From Ponds*.
<http://edis.ifas.ufl.edu/FA117>

UF/IFAS Fact Sheet FA-118 *Grading Ornamental Fish*. <http://edis.ifas.ufl.edu/FA118>

UF/IFAS Fact Sheet FA-119 *On Farm Transport of Ornamental Fish*. <http://edis.ifas.ufl.edu/FA119>

UF/IFAS Fact Sheet VM-77 *Use of Formalin to Control Fish Parasites*. <http://edis.ifas.ufl.edu/VM061>

UF/IFAS Fact Sheet VM-78 *Bath Treatment for Sick Fish*. <http://edis.ifas.ufl.edu/VM037>

UF/IFAS Fact Sheet VM-86 *Use of Salt in Aquaculture*. <http://edis.ifas.ufl.edu/VM007>

UF/IFAS Fact Sheet VM-147 *Incorporating Pet Fish Into Your Small Animal Practice*.
<http://edis.ifas.ufl.edu/VM108>

SRAC Publication No. 410 *Calculating Treatments for Ponds and Tanks*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/20/>

SRAC Publication No. 462 *Nitrite in Fish Ponds*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/25/>

SRAC Publication No. 463 *Ammonia in Fish Ponds*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/25/>

SRAC Publication No. 464 *Interactions of pH, Carbon Dioxide, Alkalinity, and Hardness in Fish Pond*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/25/>

SRAC Publication No. 468 *Carbon Dioxide in Fish Ponds*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/25/>

SRAC Publication No. 474 *The Role of Stress in Fish Disease*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/26/>

SRAC Publication No. 3900 *Anesthetics in Aquaculture*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/18/>

SRAC Publication No. 4601 *Measuring Dissolved Oxygen Concentration in Aquaculture*.
<https://srac.tamu.edu/index.cfm/event/CategoryDetails/whichcategory/25/>

References and Further Reading

Avault, J.W. 1996. *Fundamentals of Aquaculture*. Baton Rouge, LA: AVA Publishing Company Inc.

Noga, E.J. 2000. *Fish Disease: Diagnosis and Treatment*. Ames, IA: Iowa State University Press.

Wedemeyer, G.A. 1996. *Physiology of Fish in Intensive Culture Systems*. New York, NY: Chapman and Hall.

Table 4. Various-size common ornamental fish shipped in a full square-bottomed bag with 2 gallons of water

Fish Group	Small Fish	Medium Fish	Large Fish
Tetras	400/bag 1" - 1 $\frac{1}{8}$ " glow light, white clouds	300/bag 1 $\frac{1}{8}$ " - 1 $\frac{1}{2}$ " black neons, serpae, lemon, head + tail light	200-250/bag 1 $\frac{1}{2}$ " - 2 $\frac{1}{2}$ " diamonds, black, phantoms, columbians
Barbs	400/bag $\frac{5}{8}$ " - $\frac{3}{4}$ "	250/bag $\frac{3}{4}$ " - 1 $\frac{1}{2}$ " tiger	150/bag 1 $\frac{1}{2}$ " - 2"
Gouramis	NA	125-150/bag 1 $\frac{3}{4}$ " - 2"	75-100/bag 2" - 2 $\frac{1}{2}$ "
Angelfishes	150/bag $\frac{3}{4}$ " "five-cent"	75-100/bag 1" "twenty-five-cent"	20-25/bag 2" - 2 $\frac{1}{2}$ " "Silver dollar size"
Livebearers	NA	300/bag 1 $\frac{1}{2}$ " - 2" platies, mollies	250/bag 2 - 2 $\frac{1}{2}$ " swordtails